

## Review Article

# The Role of Peroxisome Proliferator Activated Receptor-gamma (PPAR $\gamma$ ) in Drug-Induced Hepatotoxicity: A Narrative Review

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## Abstract

**Background:** Drug-induced hepatotoxicity is an acute or chronic liver damage due to drugs or herbals. Its diagnosis is difficult since the presentation is similar to various hepatobiliary disorders. It is one of the major reasons for drug failures at clinical phases of drug-development and withdrawal from the market. Peroxisome proliferator-activated receptors (PPARs) are transcription factors that belong to the nuclear hormone receptor superfamily; comprising PPAR $\alpha$ , PPAR $\gamma$  and PPAR $\beta/\delta$ . They have different distribution and expression profiles, and this leads to different clinical outcomes. This review focuses on the protective role of the PPAR $\gamma$  in liver damage giving special emphasis to drug-induced hepatotoxicity.

**Methods:** This narrative review employed a systematic search strategy to identify relevant literature on the role of PPAR $\gamma$  in drug-induced hepatotoxicity. We searched major academic databases such as PubMed, Elsevier, EMBASE, Web of Science, and Google Scholar databases using a combination of keywords related to PPAR $\gamma$ , drug-induced liver injury (DILI), and hepatotoxicity. We included original research articles, reviews, and meta-analyses published in English without time bound. The retrieved articles were critically appraised for their methodology, quality of evidence, and relevance to the topic of interest.

**Result and Conclusion:** Ligands acting on PPAR $\gamma$  showed hepatoprotective activity and this protective effect is mediated through activation of Nrf2 and enhancing PPAR $\gamma$  mRNA expression. Further studies are recommended to understand the molecular mechanisms of hepatoprotective agents acting on PPAR $\gamma$ .

**Keywords:** Anti-tuberculosis drugs, Cyclophosphamide, Hepatotoxicity, Paracetamol, Peroxisome proliferator-activated receptors

## Introduction

### Drug-Induced Hepatotoxicity

Liver is one of the most important organs of humans with several functions (1). It is essential to synthesize various body proteins and to detoxify endogenous toxic metabolic byproducts and ingested toxins by the organism. Thus, loss of total liver function might lead to death within minutes (2). It involves in various biochemical pathways including growth, nutrient supply, fight against diseases, energy provision and reproduction(3). These unique involvements in metabolism and its relationship with the gastrointestinal tract made liver a target for drug-induced toxicity. After administration, drugs are generally metabolized by liver to biologically inactive forms (metabolites) and then eliminated from the body. However, certain drugs may be metabolized into metabolites that can lead to liver damage (4). Due to this reason, drug-induced liver damage became a leading cause of failures in drug development at clinical phases of investigation, and about one-fourth of drugs are prematurely terminated or withdrawn from market (5, 6).

Drug-induced damage to hepatic and bile duct cells may also result in cholestasis. In turn, cholestasis that leads to intrahepatic accumulation of excretion products and toxic bile acids. This accumulation can lead

to further hepatic injury (7, 8). Fulminant liver failure, where patients without a history of liver disease present with hepatic encephalopathy and coagulopathy preceding jaundice, is the most detrimental clinical presentation in drug-induced liver damage (9).

Two major mechanisms are involved in drug-induced hepatotoxicity. These are intrinsic and idiosyncratic hepatotoxicity (10). Intrinsic hepatotoxicity is dose-related toxicity. As exposure increases, a threshold is reached, above which individuals respond with toxicity that becomes more severe with increasing dose. This can affect all individuals at some dose and predictable using routine animal testing (11). Idiosyncratic hepatotoxicity, on the other hand, a toxicity that can be further subdivided to immunologic and metabolic toxicities. Immunologic idiosyncrasy refers to injury in the presence of clinical signs of drug hypersensitivity such as fever, rash, eosinophilia, and a prompt response to re-challenge with the drug. Conversely, metabolic idiosyncrasy refers to an injury in the absence of these clinical signs (12). The deficiency of PPAR $\gamma$  in hepatic stellate cells is associated with excessive fibrotic tissue formation in the liver (13). This review focuses on the potential role of PPAR $\gamma$  in drug-induced hepatotoxicity.

## Peroxisome proliferator-activated receptors (PPARs)

Peroxisome proliferator-activated receptors (PPARs) were discovered in 1990. They are categorized as members of the nuclear hormone receptor superfamily; function as transcription factors and modulators of gene expression (14). They are involved in regulating glucose and lipid homeostasis, inflammation, proliferation and differentiation (15). Thus, such involvement results in chronic diseases related to alterations in glucose and lipid metabolism. Some of the diseases are diabetes, obesity, non-alcoholic fatty liver disease, and atherosclerosis (16).

PPARs are expressed in several tissues, including hepatocytes, adipocytes, muscles and endothelial cells. However, the affinity depends on the isoform of PPAR and different distribution and expression profiles, and this leads to different clinical outcomes (17). PPARs are ligand-activated transcription factors and comprise three subtypes: PPAR $\alpha$ , PPAR $\gamma$ , and PPAR $\beta/\delta$  (18). When they are activated by specific ligands, all the three receptors become transcription factors that regulate the expression of distinct target genes (19). PPAR $\alpha$  is highly expressed in brown adipose tissue, which is followed by liver, heart, kidney, and skeletal muscle. Whereas, PPAR $\gamma$  is expressed mainly in adipose tissue

but also, in the heart, kidney, colon, spleen, intestine, skeletal muscle, liver, and macrophages at lower levels. The third one, PPAR $\beta/\delta$  is the least understood and it is ubiquitously expressed (20).

PPARs play a major role in the down regulation of oxidative stress, and mitochondrial and proteasomal dysfunction (21). PPAR agonists are reported to attenuate oxidative stress-mediated diseases (22). It has been shown that liver diseases mediated through oxidative stress can be ameliorated by various agents, those acting on PPARs (23-26).

### Role of PPAR $\gamma$ in Liver

PPARs exert transcriptional activity in the liver; regulating a wide spectrum of physiological functions. Some of them are regenerative mechanisms, cholesterol and bile acid homeostasis, lipid and glucose metabolism, cell differentiation/proliferation and inflammatory responses (27). In patients diagnosed with fatty liver disease, hepatic expression of PPAR $\gamma$  is involved in hepatic steatosis, insulin sensitivity, and triglyceride clearance (28). It has been reported that PPAR $\gamma$  agonists might have various therapeutic advantages including anti-diabetic activity.

Thiazolidinediones, for instance, are currently available anti-diabetic agents acting on PPAR $\gamma$ . They promote insulin sensitization and improve dyslipidemia in

diabetic patients (29,30). The first approved drug from the thiazolidinedione class was troglitazone. But, it has been withdrawn from market in 2000 due to its fatal hepatotoxicity (31). This idiosyncratic hepatotoxicity is mediated through oxidative stress and it involves mitochondrial dysfunction (32,33). The other agents, rosiglitazone and pioglitazone, have only rarely have been linked to acute liver injury. This observation could indicate that hepatotoxicity is not a class effect for thiazolidinedione. Baseline and routine monitoring of alanine aminotransferase levels as well as monitoring for clinical symptoms of liver injury, are recommended (34).

Adipose PPAR $\gamma$  deficiency elicits secondary liver phenotypes, encompassing fatty liver, enhanced gluconeogenesis, and decreased response to insulin action on hepatic glucose production. Thus, loss of fat PPAR $\gamma$  may result in progressive steatosis, lipodystrophy, and insulin resistance in the liver and fat (35). PPAR $\gamma$  co-activator 1 $\alpha$  plays an important role in the intermediary metabolism by co-activating key transcription factors of hepatic gluconeogenesis and glucose uptake in muscles (36). Furthermore, PPAR $\gamma$  may directly affect liver and pancreatic  $\beta$ -cells to improve glucose homeostasis (37).

PPAR $\gamma$  agonists decrease pro-inflammatory cytokines in macrophages. Through PPAR $\gamma$

activity, berberine inhibited the expression and production of tumor necrosis factor  $\alpha$ , monocyte chemo-attractant protein 1, and interleukin-6 in acetylated low-density lipoprotein-stimulated macrophages (38). Administration of PPAR $\gamma$  agonists, through activation of PPAR $\gamma$  signaling, protects liver against fibrosis and inflammation in mice and rats (39,40). There is a positive correlation between PPAR protein expression and total superoxide dismutase (SOD) activity. But, a negative correlation was found with the activation of NF- $\kappa$ B p65 protein expression in obstructive jaundice rats. SOD protein expression and enzyme activity will be decreased as a result of reductions in PPAR expression, which might lead to enhanced oxidative stress and lipid peroxidation in the liver (41). PPAR $\gamma$  has shown wide anti-inflammatory effects and plays a major role in controlling fibrogenesis and reducing oxidative stress. PPAR $\gamma$  ligands might be potential anti-fibrotic candidates for the management and prevention of hepatic fibrosis (42,43).

## Materials and Methods

### Literature Search

Authors performed a literature search in December 2023 from the PubMed, Elsevier, EMBASE, Web of Science and Google Scholar databases. The following terms (i) PPAR $\gamma$ /Peroxisome proliferator-activated

receptor gamma and (ii) Hepatotoxicity or Liver damage or Drug-induced liver injury or drug-induced liver disease or drug-induced liver damage were used during the search.

### **Inclusion and Exclusion Criteria**

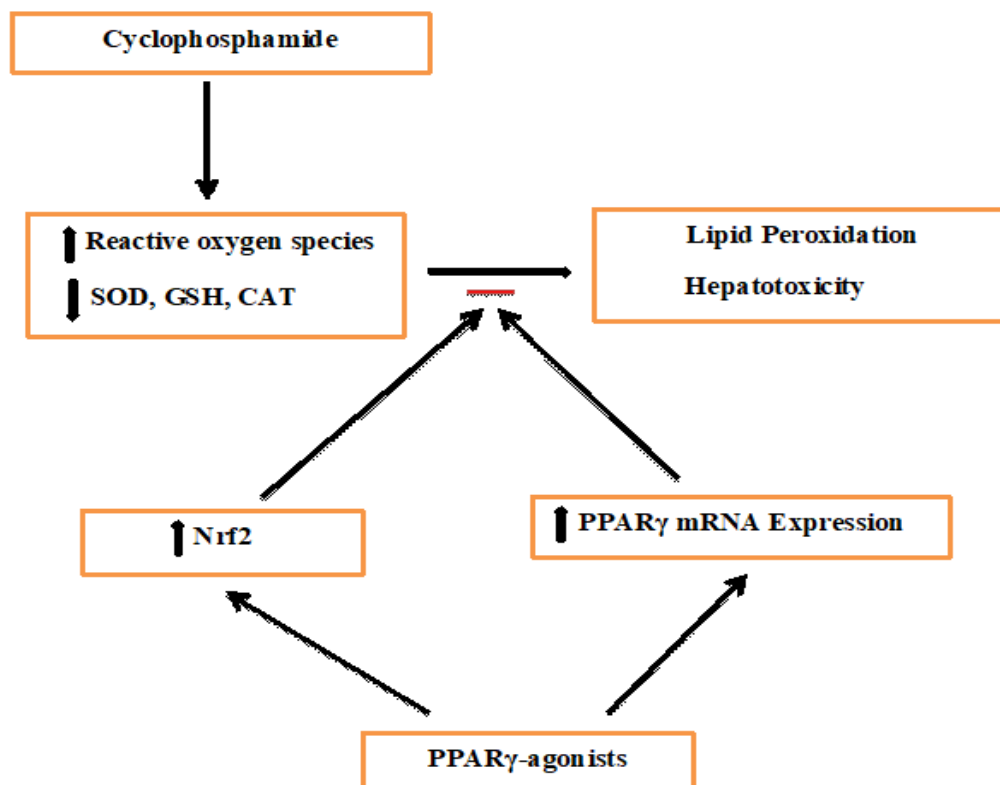
Studies published in peer-reviewed journals in English language, studies with full text, experimental studies providing data on PPAR $\gamma$  effects on drug induced hepatotoxicity were included in this study. Reviews, duplicate studies, studies without detailed research outcome were excluded.

### **PPAR $\gamma$ in Cyclophosphamide-Induced Hepatotoxicity**

Cyclophosphamide (CP) is an alkylating agent used to treat various types of cancer, lupus nephritis, and nephrotic syndrome (44). Even though CP has a broad spectrum of applications, there are deleterious effects shown in humans and experimental animals (45, 46). Since kidney and liver are the two important organs for CP metabolism and excretion, nephrotoxicity and hepatotoxicity are considered as main side effects of this drug on the abovementioned organs (47). These and other toxicities limited the clinical use of CP (48).

CP is a prodrug that needs metabolic activation by hepatic microsomal CYP<sub>450</sub> mixed function oxidase system. This activation gives phosphoramidate mustard and acrolein. During these processes, induction of oxidative stress has been noticed (49). CP is presumed to enable the production of free radicals and inhibit endogenous antioxidant activities, including SOD, glutathione (GSH) and catalase (CAT) (50).

Reactive oxygen species (ROS) lead to lipid peroxidation of the cell membrane and results in loss of cell membrane integrity. Treatment with PPAR $\gamma$ -agonists showed hepatoprotective activity through an increment of the PPAR $\gamma$  mRNA expression (51). For instance, Umbelliferone, a coumarin derivative, administration showed hepatoprotective effects through activation of nuclear factor erythroid 2-related factor 2 (Nrf2) and PPAR $\gamma$ , and subsequent suppression of inflammation and oxidative stress in CP-induced hepatotoxicity (**Figure 1**) (52). Nrf2 is a transcription factor, which is known to induce various antioxidant and cytoprotective genes (53). The following table is a summary of the evidence showing the role of PPAR $\gamma$  in CP-induced hepatotoxicity (Table 1).



**Figure 1:** Major Hepatoprotective Mechanisms of PPAR $\gamma$ -agonists, CAT (catalase), GSH, (glutathione), Nrf2 (nuclear factor erythroid 2-related factor 2), (SOD (superoxide dismutase)).

**Table 1:** The Relationship and Protective Role of PPAR $\gamma$  agonists in the Cyclophosphamide-Induced Hepatotoxicity

Ligand	Method and Intervention	Major Outcome	References
Fenofibrate (FEN) (PPAR $\alpha$ agonist) and pioglitazone (PIO) (PPAR $\gamma$ agonist)	<ul style="list-style-type: none"> <li>Rats were administered with FEN and PIO (150 and 10 mg/kg/day, respectively) for 4 weeks orally in a different group, then five days before the end of the experiment, both groups were administered cyclophosphamide (CP) (150 mg/kg, i.p.).</li> </ul>	<ul style="list-style-type: none"> <li>Pretreatment with PIO, but not FEN, before CP challenge improved hepatic function and liver histology, and significantly reversed oxidative and inflammatory parameters.</li> <li>Activation of PPAR<math>\gamma</math>, but not PPAR<math>\alpha</math>, showed protection against CP-induced hepatotoxicity, through activation of antioxidant and anti-inflammatory mechanisms.</li> <li>To conclude, PPAR<math>\gamma</math> activation might have hepatoprotective effects, as evidenced by PIO.</li> </ul>	(54)
Umbelliferone (UMB), a coumarin derivative.	<ul style="list-style-type: none"> <li>Rats were administered with UMB (50 and 100 mg/kg, orally) two weeks prior to CP injection. Five days after CP (150 mg/kg, i.p) administration, the rats were sacrificed and samples were collected for analyses.</li> <li>PPAR<math>\gamma</math> mRNA abundance was determined by quantitative reverse transcription-polymerase chain reaction (qRT-PCR).</li> </ul>	<ul style="list-style-type: none"> <li>Supplementation of UMB attenuated CP-induced inflammation and oxidative stress, as evidenced by restoration of the antioxidant defense activity and expression, and suppression of the pro-inflammatory cytokines.</li> <li>CP-induced alterations in liver histology were reduced by UMB.</li> <li>CP-induced rats showed significant down-regulation of Nrf2, HO-1 and PPAR<math>\gamma</math>, an effect that was markedly reversed by UMB.</li> <li>UMB pretreatment prevented down-regulation of PPAR<math>\gamma</math> mRNA and resulted in marked up-regulation of liver PPAR<math>\gamma</math> protein expression.</li> <li>The hepatoprotective activity of UMB might depend on co-activation of PPAR<math>\gamma</math> and Nrf2, and subsequent suppression of oxidative stress and inflammation.</li> </ul>	(52)
Hesperidin (HES), a flavonoid, is isolated from the orange <i>Citrus aurantium</i> .	<ul style="list-style-type: none"> <li>The rats were administered with a single dose of CP (200 mg/kg, i.p), followed by treatment with HES (25 and 50 mg/kg, orally) for 11 consecutive days. At the end of the experiment, blood samples and liver tissue were collected.</li> <li>Quantitative polymerase chain reaction (qPCR) was used to determine liver PPAR<math>\gamma</math>-mRNA expression level.</li> </ul>	<ul style="list-style-type: none"> <li>CP resulted in reduced glutathione content, and the activities of the antioxidant enzymes, including superoxide dismutase, catalase, and glutathione peroxidase, were reduced. CP administration induced downregulation of PPAR<math>\gamma</math> and upregulation of NF-<math>\kappa</math>B and iNOS mRNA expression.</li> <li>Administration of HES rejuvenated the altered markers in a dose-dependent manner.</li> <li>To conclude, this protective effect might be mediated through up-regulation of hepatic PPAR<math>\gamma</math> expression and abrogation of inflammation and oxidative stress.</li> </ul>	(51)

- 18 $\beta$ -Glycyrrhetic acid (18 $\beta$ -GA)
- Rats were administered 18 $\beta$ -GA (25 and 50 mg/kg, orally) for 2 weeks prior to CP (150 mg/kg, i.p) injection. Five days after CP administration, rats were sacrificed and samples were collected.
  - Gene expression analysis was performed using quantitative reverse transcription polymerase chain reaction.
  - Administration of 18 $\beta$ -GA prevented CP-induced oxidative stress and inflammation as evidenced by restoration of the antioxidant defenses and through diminishing the pro-inflammatory cytokines, lipid peroxidation, and NO production.
  - 18 $\beta$ -GA (50 mg/kg) administration produced significant up-regulation of both Nrf2 and PPAR $\gamma$  in the liver of CP-administered rats and effective than 25 mg/kg.
  - 18 $\beta$ -GA protected liver against CP-induced hepatotoxicity through activation of Nrf2 and PPAR $\gamma$ .
- (55)
- Gamma-glutamylcysteine ethyl ester (GCEE), a precursor of glutathione
- Rats were administered with GCEE (100 mg/kg) for 15 days and CP (150 mg/kg) on day 16. At day 21, rats were sacrificed and various samples were collected.
  - GCEE administration suppressed lipid peroxidation and nitric oxide production and restored glutathione, and enzymatic antioxidants in the liver, which were associated with downregulation of COX-2, iNOS, and NF- $\kappa$ B.
  - GCEE up-regulated PPAR $\gamma$  and imparted its hepatoprotective effect through activating PPAR $\gamma$ , preventing GSH depletion, and attenuating oxidative stress, inflammation, and apoptosis.
  - Quantitative RT-PCR was used to study gene expression
- (56)

Key: Cyclophosphamide (CP), nuclear factor erythroid 2-related factor 2 (Nrf2), heme oxygenase (HO-1), nuclear factor-kappa B (NF- $\kappa$ B), inducible nitric oxide synthase (iNOS),

## Role of PPAR $\gamma$ in Other Drug-Induced Hepatotoxicities

Paracetamol (PCM) is an analgesic and antipyretic drug that is safe at its therapeutic doses. However, it is associated with acute liver damage during acute overdose (57). PCM-induced hepatotoxicity is one of the most common causes of liver failure (58). PPAR $\gamma$  may function to protect mitochondria from ROS, resulting from PCM overdose (59).

In a recent study, both curcumin and silymarin showed anti-apoptotic and anti-inflammatory effects on liver tissues by decreasing Bax and increasing Bcl2 mRNA expression in PCM-induced hepatotoxicity in adult male albino rats. PPAR $\gamma$  mRNA expression has been reported to significantly increase with curcumin treatment. The possible hepatoprotective mechanism could be via activating PPAR $\gamma$ , which is involved in the inhibition of sterol regulatory element binding protein 2 (SREBP-2) and low-density lipoprotein receptor (LDLR) gene expression. These proteins could reinstate lipid storage capacity of hepatic stellate cells and protect against liver steatosis and fibrosis (57).

Some anti-tuberculosis drugs are associated with hepatotoxicity, including isoniazid, pyrazinamide and rifampicin (60, 61). Isoniazid is a highly effective drug in the management of tuberculosis (62). Even

though the biochemical basis of isoniazid-induced hepatotoxicity remains elusive, it has been reported that isoniazid can disrupt multiple endobiotics metabolic pathways, which are all critical for hepatocellular functions (63). Hydrazine, the reactive metabolite of isoniazid, was shown to inhibit the activity of solubilized complex I in mouse hepatocytes resulting in mitochondrial oxidative stress. Thus, isoniazid-induced oxidative stress affects mitochondrial dynamics and might lead to liver damage (64).

As reported by *Mahmoud* and his colleagues, administration of berberine (25 and 50 mg/kg, orally) for 45 days showed a protective effect against isoniazid-induced oxidative stress and inflammation in rats, through up-regulating PPAR $\gamma$  and subsequently suppressing NF- $\kappa$ B, iNOS and release of the pro-inflammatory cytokines. Gene expression analysis using quantitative polymerase chain reaction in rats treated with berberine showed increased liver PPAR $\gamma$  mRNA expression when compared with the control groups (13).

Chronic alcohol intake has also been reported to cause alcoholic liver disease which involves three stages; steatosis (fatty liver), hepatitis, and cirrhosis. The main pathologic mechanisms for fatty liver development include an increase in plasma corticosterone, induction of zinc deficiency, hyperlipolysis

of adipose tissue and reverse triglyceride (TG) transport to liver, and induction of leptin deficiency (65). Treatment with 4-Methylcoumarin-(5,6-g)-hesperetin (4-MCH) was found to attenuate inflammatory responses in alcoholic hepatitis through PPAR $\gamma$  activation in mice. In addition, 4-MCH was found to suppress the release of inflammatory cytokines including interleukin-6 (IL-6) and tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ). The expression of PPAR $\gamma$  was also up-regulated in the 4-MCH group (66).

High intake of the western diets such red meat and processed meat, high-fat dairy products and sweet items, is associated with obesity and metabolic syndrome. This also increases the risk of type 2 diabetes mellitus and cardiovascular diseases (67). Animals fed with a western diet develop advanced nonalcoholic fatty liver disease, including nonalcoholic steatohepatitis (NASH) and hepatic fibrosis (68). Chronic dietary intake of quercetin is reported to reduce oxidative stress. Literature reports showed that Quercetin can alleviate hepatic fat accumulation in mice liver fed with western diet. This might be through increasing PPAR $\gamma$  expression (67).

## Conclusion

The peroxisome proliferator-activated receptors are members of the nuclear receptor superfamily of ligand-dependent transcription factors. They are involved in regulating glucose and lipid homeostasis, inflammation, proliferation and differentiation. Thus, they play a major role in the down-regulation of oxidative stress, mitochondrial and proteasomal dysfunction. Even though the exact mechanism is still obscure, various agonists of PPAR $\gamma$  showed a promising effect in preventing drug-induced hepatotoxicity through activation of Nrf2 and enhancing PPAR $\gamma$  mRNA expression. Since hepatotoxicity is a major concern in drug development, further researches including pre-clinical and clinical studies should be encouraged on hepatoprotective agents acting through PPAR $\gamma$ .

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## Consent for publication

All authors provided consent for the publication of the manuscript detailed above.

## Author Contributions

All authors participated in the conception of the work, data collection, drafting the article, critical revision of the article and in the final approval of the version to be published.

## References

1. Vaja R, Rana M. Drugs and the liver. *Anaesthesia Intensive Care Med.* 2020;21(10):517-23.
2. Ozougwu JC. Physiology of the liver. *Int J Res Pharm Biosci.* 2017;4(8):13-24.
3. Pandit A, Sachdeva T, Bafna P. Drug-induced hepatotoxicity: a review. *J Appl Pharm Sci.* 2012;2(5):233-43.
4. Srivastava A, Maggs JL, Antoine DJ, Williams DP, Smith DA, Park BK. Role of Reactive Metabolites in Drug-Induced Hepatotoxicity. In: Uetrecht J, editor. *Adverse Drug Reactions.* Berlin, Heidelberg: Springer Berlin Heidelberg; 2010. p. 165-94.
5. Low Y, Uehara T, Minowa Y, Yamada H, Ohno Y, Urushidani T, Alexander S, Eugene M, Viktor K, Denis F, Hao Z, Ivan R, Alexander. Predicting drug-induced hepatotoxicity using QSAR and toxicogenomics approaches. *Chem Res Toxicol.* 2011;24(8):1251-62.
6. Babai S, Auclert L, Le-Louet H. Safety data and withdrawal of hepatotoxic drugs. *Therap.* 2021;76(6):715-23.
7. Jaeschke H, Gores GJ, Cederbaum AI, Hinson JA, Pessayre D, Lemasters JJ. Mechanisms of hepatotoxicity. *Toxicol Sci.* 2002;65(2):166-76.
8. Shehu AI, Ma X, Venkataramanan R. Mechanisms of drug-induced hepatotoxicity. *Clin Liver Disease.* 2017;21(1):35-54.
9. Yoon E, Babar A, Choudhary M, Kutner M, Pysopoulos N. Acetaminophen-induced hepatotoxicity: a comprehensive update. *J Clinical Translational Hepatology.* 2016;4(2):131.
10. Zhang X, Ouyang J, Thung SN. Histopathologic Manifestations of Drug-induced Hepatotoxicity. *Clinics in Liver Disease.* 2013;17(4):547-64.
11. Roth RA, Ganey PE. Intrinsic versus idiosyncratic drug-induced hepatotoxicity-two villains or one? *J Pharmacol Experimental Therapeut.* 2010;332(3):692-7.
12. Walgren JL, Mitchell MD, Thompson DC. Role of Metabolism in Drug-Induced Idiosyncratic Hepatotoxicity. *Critical Rev Toxicol.* 2005;35(4):325-61.

13. Mahmoud AM, Germoush MO, Soliman AS. Berberine attenuates isoniazid-induced hepatotoxicity by modulating peroxisome proliferator-activated receptor gamma, oxidative stress and inflammation. *Int J Pharmacol.* 2014;10(8):451-60.
14. Youssef J, Badr M. Peroxisome proliferator-activated receptors and cancer: challenges and opportunities. *British J Pharmacol.* 2011;164(1):68-82.
15. Peters JM, Shah YM, Gonzalez FJ. The role of peroxisome proliferator-activated receptors in carcinogenesis and chemoprevention. *Nature Rev Cancer.* 2012;12(3):181-95.
16. Yessoufou A, Wahli W. Multifaceted roles of peroxisome proliferator-activated receptors (PPARs) at the cellular and whole organism levels. *Swiss medical weekly.* 2010;140.
17. Grygiel GB. Peroxisome proliferator-activated receptors and their ligands: nutritional and clinical implications - a review. *Nutrition J.* 2014;13(1):17.
18. Tyagi S, Gupta P, Saini AS, Kaushal C, Sharma S. The peroxisome proliferator-activated receptor: A family of nuclear receptors role in various diseases. *J Advanced Pharmaceut Technol Res.* 2011;2(4):236.
19. Rosenson RS, Wright RS, Farkouh M, Plutzky J. Modulating peroxisome proliferator-activated receptors for therapeutic benefit? Biology, clinical experience, and future prospects. *Am Heart J.* 2012;164(5):672-80.
20. Pirat C, Farce A, Lebègue N, Renault N, Furman C, Millet R, Saïd Y, Silvia S, Pascal B, Pierre D, Philippe C. Targeting Peroxisome Proliferator-Activated Receptors (PPARs): Development of Modulators. *J Med Chem.* 2012;55(9):4027-61.
21. Agarwal S, Yadav A, Chaturvedi RK. Peroxisome proliferator-activated receptors (PPARs) as therapeutic target in neurodegenerative disorders. *Biochem Biophys Res Commun.* 2017;483(4):1166-77.
22. Yu X, Shao X, Sun H, Li Y, Yang J, Deng Y, Yuan-Gui H. Activation of cerebral peroxisome proliferator-activated receptors gamma exerts neuroprotection by inhibiting oxidative stress following pilocarpine-induced status epilepticus. *Brain Res.* 2008;1200:146-58.
23. Coleman JD, Prabhu KS, Thompson JT, Reddy PS, Peters JM, Peterson BR, Channa R, John PV. The oxidative stress mediator 4-hydroxynonenal is an intracellular agonist of the nuclear receptor

- peroxisome proliferator-activated receptor- $\beta/\delta$  (PPAR $\beta/\delta$ ). *Free Radical Biol Med.* 2007;42(8):1155-64.
24. Riserus U, Sprecher D, Johnson T, Olson E, Hirschberg S, Liu A, Zeke F, Priti H, Duncan R, Leli SB, Jay CS, Samar B, Jane C, Barbara AF, Sandy MH, Theodore D, Niall RM, Peter M, Stephen O, Pauline S, Tim W, David H, Keith NF, Fredrik K. Activation of Peroxisome Proliferator-Activated Receptor (PPAR) $\delta$  promotes reversal of multiple metabolic abnormalities, reduces oxidative stress, and increases fatty acid oxidation in moderately obese men. *Diabetes.* 2008;57(2):332-9.
25. Li JL, Wang QY, Luan HY, Kang ZC, Wang CB. Effects of L-carnitine against oxidative stress in human hepatocytes: involvement of peroxisome proliferator-activated receptor alpha. *J Biomed Sci.* 2012;19(1):32.
26. Sung B, Park S, Yu BP, Chung HY. Amelioration of age-related inflammation and oxidative stress by PPAR $\gamma$  activator: Suppression of NF- $\kappa$ B by 2,4-thiazolidinedione. *Experimental Gerontolog.* 2006;41(6):590-9.
27. Peyrou M, Ramadori P, Bourgoin L, Foti M. PPARs in Liver Diseases and Cancer: Epigenetic Regulation by MicroRNAs. *PPAR Res.* 2012;2012:757803.
28. Rey JW, Noetel A, Hardt A, Canbay A, Alakus H, Zur Hausen A, Hans PD, Uta D, Margarete O. Pro12Ala polymorphism of the peroxisome proliferator-activated receptor  $\gamma$ 2 in patients with fatty liver diseases. *World J Gastroenterol.* 2010;16(46):5830-7.
29. Duan SZ, Usher MG, Mortensen RM. Peroxisome proliferator-activated receptor-gamma-mediated effects in the vasculature. *Circ Res.* 2008;102(3):283-94.
30. Fogo AB. Potential for peroxisome proliferator-activated receptor-gamma agonists in progression: beyond metabolism. *Curr Opin Nephrol Hypertens.* 2008;17(3):282-5.
31. Wang L, Waltenberger B, Pferschy-Wenzig EM, Blunder M, Liu X, Malainer C, Tina B, Stefan S, Judith MR, Elke HH, Daniela S, Brigitte K, Rudolf B, Hermann S, Verena MD, Atanas GA. Natural product agonists of peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ): a review. *Biochem Pharmacol.* 2014;92(1):73-89.
32. Julie NL, Julie IM, Kende AI, Wilson GL. Mitochondrial dysfunction and

- delayed hepatotoxicity: another lesson from troglitazone. *Diabetologia*. 2008;51(11):2108-16.
33. Yokoi T. Troglitazone. In: Uetrecht J, editor. *Adverse Drug Reactions*. Berlin, Heidelberg: Springer Berlin Heidelberg; 2010. p. 419-35.
34. Eggleton JS, Jialal I. Thiazolidinediones. *StatPearls* [Internet]: StatPearls Publishing; 2019.
35. He W, Barak Y, Hevener A, Olson P, Liao D, Le J, Michael N, Estelita O, Jerrold MO, Ronald ME. Adipose-specific peroxisome proliferator-activated receptor  $\gamma$  knockout causes insulin resistance in fat and liver but not in muscle. *Proceed Nat Acad Sci*. 2003;100(26):15712-7.
36. Oberkofler H, Schraml E, Krempler F, Patsch W. Potentiation of liver X receptor transcriptional activity by peroxisome-proliferator-activated receptor gamma co-activator 1 alpha. *Biochem J*. 2003;371(1):89-96.
37. Kim HI, Ahn YH. Role of Peroxisome Proliferator-Activated Receptor- $\gamma$  in the glucose-sensing apparatus of liver and  $\beta$ -Cells. *Diabetes*. 2004;53(suppl 1):S60-S5.
38. Chen FL, Yang ZH, Liu Y, Li L, Liang WC, Wang XC, Zhou WB, Yang YH, Ren-Ming H. Berberine inhibits the expression of TNF $\alpha$ , MCP-1, and IL-6 in AcLDL-stimulated macrophages through PPAR $\gamma$  pathway. *Endocrine*. 2008;33(3):331-7.
39. Tahan V, Eren F, Avsar E, Yavuz D, Yuksel M, Emekli E, Nese I, Cigdem C, Hafize U, Goncagul H, Nurdan T. Rosiglitazone attenuates liver inflammation in a rat model of nonalcoholic steatohepatitis. *Digest Disease Sci*. 2007;52(12):3465-72.
40. Nan YM, Fu N, Wu WJ, Liang BL, Wang RQ, Zhao SX, Jing-Min Z, Jun Y. Rosiglitazone prevents nutritional fibrosis and steatohepatitis in mice. *Scand J Gastroenterology*. 2009;44(3):358-65.
41. Gong P, Xu H, Zhang J, Wang Z. PPAR expression and its association with SOD and NF-kB in rats with obstructive jaundice. *Biomed Res*. 2012;23(4).
42. Sun K, Wang Q, Huang X. PPAR gamma inhibits growth of rat hepatic stellate cells and TGF beta-induced connective tissue growth factor expression 1. *Acta Pharmacol Sinica*. 2006;27(6):715-23.
43. Lutchman G, Modi A, Kleiner DE, Promrat K, Heller T, Ghany M, Brian B, Rohit L, Jake L, Ahalya P, Jay H. The effects of discontinuing

- pioglitazone in patients with nonalcoholic steatohepatitis. *Hepatology*. 2007;46(2):424-9.
44. Subramaniam SR, Cader RA, Mohd R, Yen KW, Ghafor HA. Low-dose cyclophosphamide-induced acute hepatotoxicity. *Am J Case Reports*. 2013;14:345.
45. Singh C, Prakash C, Tiwari KN, Mishra SK, Kumar V. *Premna integrifolia* ameliorates cyclophosphamide-induced hepatotoxicity by modulation of oxidative stress and apoptosis. *Biomed Pharmacother*. 2018;107:634-43.
46. Ogino MH, Tadi P. Cyclophosphamide. *StatPearls* [Internet]: StatPearls Publishing; 2020.
47. Temel Y, Kucukler S, Yildirim S, Caglayan C, Kandemir FM. Protective effect of chrysin on cyclophosphamide-induced hepatotoxicity and nephrotoxicity via the inhibition of oxidative stress, inflammation, and apoptosis. *Naunyn-Schmiedeberg's Archive Pharmacol*. 2020;393(3):325-37.
48. Refaie MMM, Shehata S, El-Hussieny M, Abdelraheem WM, Bayoumi AMA. Role of ATP-Sensitive Potassium Channel (KATP) and eNOS in mediating the protective effect of Nicorandil in cyclophosphamide-induced cardiotoxicity. *Cardiovascular Toxicol*. 2020;20(1):71-81.
49. Zarei M, Shivanandappa T. Amelioration of Cyp-induced hepatotoxicity by the root extract of *Decalepis hamiltonii* in mice. *Food Chem Toxicol*. 2013;57:179-84.
50. Hamzeh M, Hosseinimehr SJ, Khalatbary AR, Mohammadi HR, Dashti A, Amiri FT. Atorvastatin mitigates cyclophosphamide-induced hepatotoxicity via suppression of oxidative stress and apoptosis in rat model. *Res Pharmaceut Sci*. 2018;13(5):440.
51. Mahmoud AM. Hesperidin protects against cyclophosphamide-induced hepatotoxicity by upregulation of PPAR $\gamma$  and abrogation of oxidative stress and inflammation. *Canadian J Physiology Pharmacol*. 2014;92(9):717-24.
52. Mahmoud AM, Germoush MO, Alotaibi MF, Hussein OE. Possible involvement of Nrf2 and PPAR $\gamma$  up-regulation in the protective effect of umbelliferone against cyclophosphamide-induced hepatotoxicity. *Biomed Pharmacother*. 2017;86:297-306.

53. Reisman SA, Aleksunes LM, Klaassen CD. Oleanolic acid activates Nrf2 and protects from acetaminophen hepatotoxicity via Nrf2-dependent and Nrf2-independent processes. *Biochem Pharmacol.* 2009;77(7):1273-82.
54. El-Sheikh AA, Rifaai RA. Peroxisome proliferator activator receptor (PPAR)- $\gamma$  ligand, but not PPAR- $\alpha$ , ameliorates cyclophosphamide-induced oxidative stress and inflammation in rat liver. *PPAR Res.* 2014;2014.
55. Mahmoud AM, Al Dera HS. 18 $\beta$ -Glycyrrhetic acid exerts protective effects against cyclophosphamide-induced hepatotoxicity: potential role of PPAR $\gamma$  and Nrf2 upregulation. *Gene Nutrition.* 2015;10(6):41.
56. Alqahtani S, Mahmoud AM. Gamma-Glutamylcysteine ethyl ester protects against cyclophosphamide-induced liver injury and hematologic alterations via upregulation of PPAR $\gamma$  and attenuation of oxidative stress, inflammation, and apoptosis. *Oxid Med Cell Longev.* 2016;2016:4016209.
57. Ahmad MM, Rezk NA, Fawzy A, Sabry M. Protective effects of curcumin and silymarin against paracetamol induced hepatotoxicity in adult male albino rats. *Gene.* 2019;712:143966.
58. Ye D, Wang Y, Li H, Jia W, Man K, Lo CM, Yu W, Karen SL, Aimin X. Fibroblast growth factor 21 protects against acetaminophen-induced hepatotoxicity by potentiating peroxisome proliferator-activated receptor coactivator protein-1 $\alpha$ -mediated antioxidant capacity in mice. *Hepatology.* 2014;60(3):977-89.
59. Patterson AD, Shah YM, Matsubara T, Krausz KW, Gonzalez FJ. Peroxisome proliferator-activated receptor alpha induction of uncoupling protein 2 protects against acetaminophen-induced liver toxicity. *Hepatology.* 2012;56(1):281-90.
60. Yew W, Leung C. Antituberculosis drugs and hepatotoxicity. *Respirolog.* 2006;11(6):699-707.
61. John P, Kale PP. Prominence of oxidative stress in the management of anti-tuberculosis drugs related hepatotoxicity. *Drug Metabolism Lett.* 2019;13(2):95-101.
62. Wang P, Pradhan K, Zhong X-b, Ma X. Isoniazid metabolism and hepatotoxicity. *Acta Pharmaceut Sinica B.* 2016;6(5):384-92.
63. Wang P, Li F, Lu J, Grant DM, Zhong X-b, Ma X. Deficiency of N-

- Acetyltransferase potentiates isoniazid-endobiotics interactions and contributes to isoniazid hepatotoxicity. *The FASEB J.* 2018;32(1\_supplement):lb654-lb.
64. Ramachandran A, Visschers RG, Duan L, Akakpo JY, Jaeschke H. Mitochondrial dysfunction as a mechanism of drug-induced hepatotoxicity: current understanding and future perspectives. *J Clinical Ttrans Res.* 2018;4(1):75.
65. Zhang W, Sun Q, Zhong W, Sun X, Zhou Z. Hepatic peroxisome proliferator-activated receptor gamma signaling contributes to alcohol-induced hepatic steatosis and inflammation in mice. *Alcoholism: Clin Exp Res.* 2016;40(5):988-99.
66. Meng H-W, You H-M, Yang Y, Zhang Y-L, Meng XM, Ma TT, Cheng H, Jun L. 4-Methylcoumarin-[5,6-g]-hesperetin attenuates inflammatory responses in alcoholic hepatitis through PPAR- $\gamma$  activation. *Toxicol.* 2019;421:9-21.
67. Kobori M, Masumoto S, Akimoto Y, Oike H. Chronic dietary intake of quercetin alleviates hepatic fat accumulation associated with consumption of a Western-style diet in C57/BL6J mice. *Mol Nutr Food Res.* 2011;55(4):530-40.
68. Yang P, Wang Y, Tang W, Sun W, Ma Y, Lin S, Jia J, Long J, Hang S, Zhiyuan S, Liqing Y. Western diet induces severe nonalcoholic steatohepatitis, ductular reaction, and hepatic fibrosis in liver CGI-58 knockout mice. *Sci Reports.* 2020;10(1):1-13.